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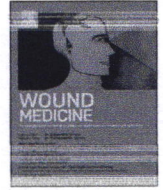
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## Original research article

## Effects of platelet-rich plasma and carbonated hydroxyapatite combination on cranial defect Bone Regeneration: An animal study



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## ABSTRACT

**Background:** Recently, platelet-rich plasma (PRP) has become popular in the tissue engineering field. PRP has a high concentration of platelets that is three to five times above that of normal plasma and contains several growth factors such as platelet-derived growth factor (PDGF), platelet-derived angiogenesis factor (PDAF), platelet-derived endothelial growth factor (PDEGF), transforming growth factor-beta (TGF-β), insulin-like growth factor (IGF), fibroblast growth factor (FGF), and vascular endothelial growth factor (VEGF). Standard reconstruction of cranial bone defects involves the use of auto- or allogenic biomaterial, such as carbonated hydroxyapatite (CHA) as a scaffold for the osteogenesis process. The purpose of this study is to investigate whether any additional compound may improve the bone healing process.

**Methods:** This study involved animal experiments on white male rats (*Rattus norvegicus*). Three millimetre diameter were created in rat cranium. Samples were divided into three groups: first group, the cranial defect was grafted with CHA combined with PRP, second group, with CHA alone, and third group, the defect was left as secondary healing wound (control group). The wound healing process was observed for the presence of inflammatory cells and the occurrence of woven bone and lamellar bone. The results among the groups were compared and analysed by the Mann Whitney test using SPSS Statistics Program Package Version 22.0.

**Results:** The experimental group of 2 weeks showed no difference between inflammatory response ( $p = 0.119$ ), woven bone ( $p = 0.094$ ) and lamellar bone ( $p = 0.130$ ). At 4 weeks, a combination of PRP and CHA showed a superior growth of lamellar bone compared to CHA ( $p = 0.008$ ).

**Conclusion:** A combination of PRP and CHA as a bone regeneration scaffold showed a histologically increased bone formation.

## 1. Introduction

Platelet-rich plasma (PRP) application has emerged as a new approach to tissue regeneration and tissue engineering paradigm as a natural source of growth factors that may accelerate bone regeneration [1]. PRP acts as a biomaterial accelerator of bone formation and contains proteins and many growth factors (GF) with osteoinductive characteristics [2]. PRP has a high concentration of plasma platelets which is five times higher than that of the blood (normally 150,000–350,000 cells/ $\mu$ L) and contains at least seven growth factors such as platelet-derived growth factor (PDGF), platelet-derived

angiogenesis factor (PDAF), platelet-derived endothelial growth factor (PDEGF), transforming growth factor-beta (TGF-β), insulin-like growth factor (IGF), fibroblast growth factor (FGF), and vascular endothelial growth factor (VEGF) [3]. These growth factors are present within granules inside the concentrated platelets of PRP [4]. Growth factors are proteins that enable different cellular processes involved in tissue healing, such as infiltration, growth, differentiation, migration, cell metabolism and apoptosis. The modification of growth factors may reinforce tissue restoration by upregulating the aforementioned processes [5]. The application of PRP to autogenous bone grafts increases bone mineral density and accelerates bone regeneration and soft tissue

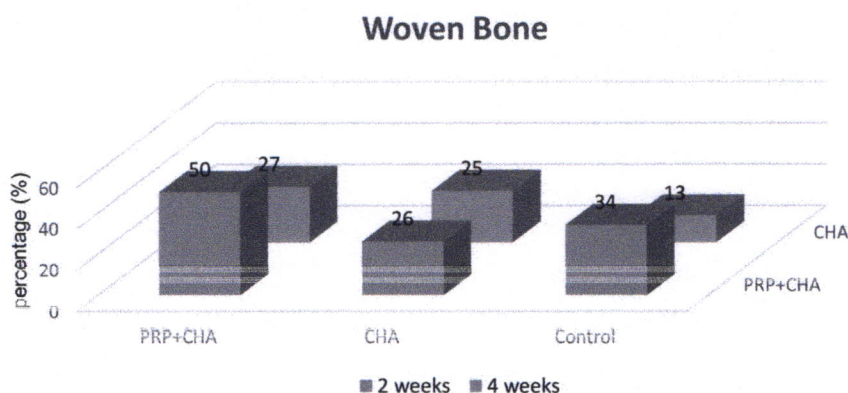
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**Table 1**  
Inflammatory response among the three experimental groups.

	2 Weeks				4 Weeks			
	Minimum	Maximum	Median	Mean	Minimum	Maximum	Median	Mean
PRP + CHA	1	4	4	3	2	3	2.5	3
CHA	4	4	4	4	3	4	4	3.8
Control	4	4	4	4	3	4	3	3.2

PRP: platelet-rich plasma, CHA: carbonate-substituted hydroxyapatite.



**Fig. 1.** Percentage of woven bone occupying the cranial defect. PRP: platelet-rich plasma, CHA: carbonated hydroxyapatite.

healing. PRP has been successfully used for mandibular reconstructions and dental implant procedures [6].

Bone healing and regeneration is one of the most important processes in craniofacial surgery. Carbonate-substituted hydroxyapatite (CHA) is a major inorganic component of natural bone, and owing to its bioactive, biodegradable and osteoconductive properties, CHA has been used extensively in biomedical implant and bone regeneration applications. CHA is also known to be biocompatible, non-toxic, non-inflammatory, and non-immunogenic, and is capable of forming a direct chemical bond with surrounding hard tissues. CHA is synthesized from calcium nitrate tetrahydrate, diammonium hydrogen phosphate and sodium hydrogen carbonate. CHA is also more osteoconductive and more resorbable than hydroxyapatite (HA) [7]. Carbonated HA contains 3–5% carbonate ions by substitution in the hydroxyapatite lattice structure and is the major mineral constituent of bone [8]. Several studies such as Ellies et al. [9] and Patel et al. [10] have reported improvement of bone formation with CHA implants compared with HA controls.

The present study aimed to evaluate the effects of PRP and CHA on bone regeneration in experimental rat cranial defects.

## 2. Methods

The study has been approved by scientific study ethical committee of Universitas Sam Ratulangi. The present study was an animal experimental study using 36 male rats (*Rattus norvegicus*) aged 20–22 weeks old and with body weight of 350–400 g. The animals were kept individually in quarantined rooms under 12-h light/dark cycle with controlled temperature and humidity, and minimal noise, and they had full access to standard dry food and water ad libitum. Each experimental group consisted of six rats. The rats were anaesthetized with an intramuscular injection of ketamine 10% (30 mg/kg body weight), and their head was then shaved over the cranium and draped in a sterile manner. Following a nasofrontal-external protuberance occipital approach, the periosteum was dissected and identical full-thickness bony defects were created with a 3-mm round bur. The defects were filled with the combination of PRP and CHA (group 1), CHA alone (group 2), and kept unfilled to serve as a control (group 3). The wounds were

closed with a non-absorbable 5/0 suture.

### 2.1. PRP preparation

A total of 3 mL autologous blood was drawn from each rat and combined with the anticoagulant citrate dextrose phosphate to prevent coagulation. The blood sample was then centrifuged at 160g for 20 min to separate the plasma containing the platelets from the red blood cells. The plasma was extracted from the top and centrifuged for an additional 15 min at 400g to separate the platelets. The platelet-poor plasma was separated from the PRP along with the buffy coat. The rats were euthanised with an overdose of diethyl ether at 2 and 4 weeks. The entire cranium was removed with a reciprocating saw. Specimens were treated with 20% formic acid decalcifying solution for three days, dehydrated with alcohol and then embedded in paraffin. The specimens were prepared routinely with haematoxylin and eosin staining. Histologic evaluation was performed at 10 times magnification.

### 2.2. Outcomes

After two weeks of treatment, the wound healing process was examined for the inflammatory response as follows: < 5 leucocyte/field of view (grade 1); 5–10 leucocyte/field of view (grade 2); 10–50 leucocyte/field of view (grade 3); > 50 leucocyte/field of view (grade 4) at 2nd and 4th weeks. The other outcome measurement was bone formation (woven bone and lamellar bone; percentage occupying the defect), and the value were compared between the groups at 2nd and 4th weeks.

### 2.3. Statistical analysis

Data were analysed using the Statistical Package for Social Sciences (SPSS) version 22.0. One-way analysis of variance (ANOVA) and Kruskal Wallis test were performed for statistical analysis. A *p* value of less than 0.05 was considered statistically significant.

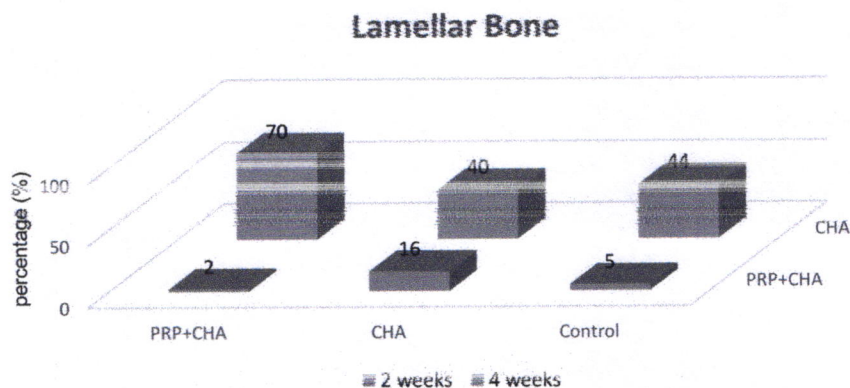


Fig. 2. Distribution data of lamellar bone regeneration. PRP: platelet-rich plasma, CHA: carbonated hydroxyapatite.

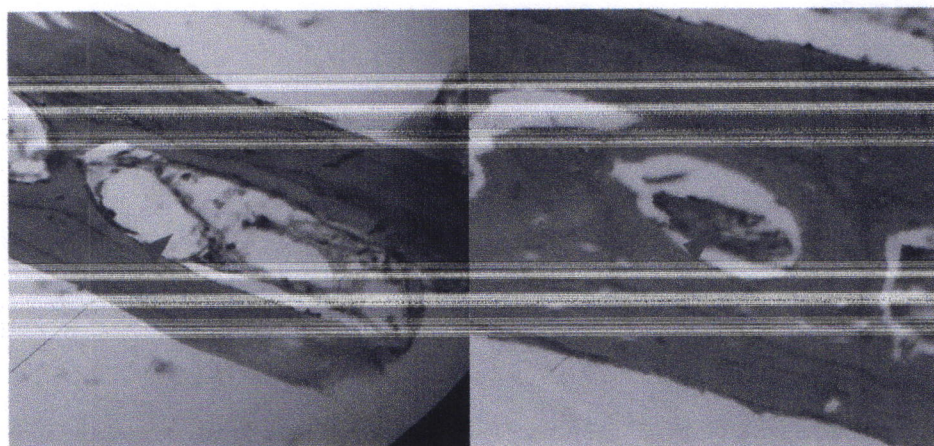


Fig. 3. Comparison of pathological features at Platelet-rich plasma (PRP) + carbonated hydroxyapatite (CHA) group after 2 weeks (left) and 4 weeks (right). At 2 weeks, minimal bone formation (empty lamellae) was observed (left arrow) with a high extent of inflammatory response and woven bone. At 4 weeks, regular parallel alignment of collagen with bone formation inside the lamellae was observed (right arrow).

### 3. Results

#### 3.1. Inflammatory response

After 2 weeks of treatment, there were no significant differences among all experimental groups, but after 4 weeks, statistically significant differences ( $p = 0.005$ ) in inflammatory cells were observed among the groups, with the lowest inflammation response observed in group 1 (PRP + CHA) (see Table 1).

#### 3.2. Bone formation

Within 2 weeks, the woven bone started to occupy the cranial defect, and there were statistically significant difference among the groups; the highest value was observed in the PRP + CHA group with 50% percentage, followed by 26% in the CHA group, and 34% in the control group (see Fig. 1). After 4 weeks, the average percentage of woven bone was not statistically different with 27% and 25% in the PRP + CHA and CHA alone groups compared with 13% in the control group (see Fig. 2).

For the lamellar bone formation, there were no significant differences between the 3 groups in 2 weeks of observation, but after 4 weeks, there was a significant ( $p = 0.009$ ) increase in lamellar bone regeneration up to 70% in the PRP + CHA group compared to 40% in the CHA alone group and 44% in the control group (see Fig. 2).

In pathological examination, after 2 weeks, a high extent of inflammatory response and woven bone was observed and lamellar bone formation was still minimal. At 4 weeks, parallel alignment of collagen into lamellae was observed and the number of inflammatory cells had decreased (see Fig. 3)

### 4. Discussion

There are four important properties in bone regeneration: [1] osteoconductive matrix which acts as a scaffold or framework into which bone growth occurs; [2] osteoinductive factors which are growth factors such as bone morphogenetic protein and transforming growth factor- $\beta$  that stimulate bone formation; [3] osteogenic cells which include primitive mesenchymal cells, osteoblasts and osteocytes; and [4] structural integrity. [11]. On the basis of this paradigm, the present study tried to boost at least 2 of the above 4 factors by using CHA as a scaffold of bone growth and PRP to fasten the production of osteoinductive factors.

During the acute phase, macro and microvascular disruption leads to hematoma formation. A serofibrinous exudate permeates the injured region with the release of vasoactive angiogenic factor from the necrotic bone and vascular tissue. Vasodilatation and hyperemia of soft tissue occurs. Bone necrosis may occur after 7–10 days if all bone marrow having cellular necrosis. Within the first 2 days, although it may evident as early 8–16 h with average maximum response at 32 h after injury, polymorphic population of pluripotent mesenchymal cells emerged. This cellular population includes macrophages, fibroblasts and other cells. Later on, the immature bone or woven bone may be seen after 12 days and became mature bone or lamellar bone after more than 4 weeks [12].

In this study, the inflammatory reaction after 2 weeks was still high as represented by more than 50 inflammatory cells in the wound cavity at all groups. There are slightly decrease for group of combination platelet-rich plasma and carbonated hydroxyapatite as 10–50 inflammatory cells. The inflammatory reaction continue to 4 weeks with average 10–50 inflammatory cells. Histologic assessment revealed that PRP and CHA are biocompatible, osteoconductive grafting material, but

inflammatory response was still observed along with their application.

The amount of immature bone tended to increase in the PRP + CHA group compared to that in the other groups in weeks 2 and 4. In week 4, the amount of mature bone was significantly increased in the PRP + CHA group compared to that in the CHA alone group. The possible reason for this condition may be the high concentration of growth factors in PRP which promote new bone formation. Other authors have claimed that there are no benefits from using PRP in bone regeneration. For instance, Schlegel et al. [13] reported that PRP does not affect bone regeneration, and Aghaloo et al. [14] observed reduced bone formation and mineral density in rabbit cranial defects treated with PRP. The results of the present study are in accordance with those of Yin et al. [4], Pryor et al. [15] and Brogгинi et al. [16].

## 5. Conclusion

The combination of PRP and CHA increased bone regeneration in experimental rat cranial defects.

## Conflict of interest declaration

We have no conflict of interest to declare.

## References

- [1] Z. Jiang qing, Liu huan ye, Zhang li ping, Wu zhi qiang, Shang de zhi, Repair of calvarial defects in rabbits with platelet-rich plasma as the scaffold for carrying bone marrow stromal cells. *Oral. Surg. Oral Med. Oral Pathol. Oral Radiol.* 113 (3) (2012) 327–333.
- [2] S. Approaches, G.F. Delivery, R.M. Edited, S. Panseri, F. Taraballi, C. Canha, Biomimetic Approaches for Tissue Healing Strategic Approaches to Growth Factors Delivery for Regenerative Medicine, (2015).
- [3] J. Alsousou, M. Thompson, P. Hulley, A. Noble, K. Willett, The biology of platelet-rich plasma and its application in trauma and orthopaedic surgery: a review of the literature, *J. Bone Joint Surg. Br.* 91 (8) (2009) 987–996.
- [4] W. Yin, C. Zhang, X. Qi, Y. Zhang, J. Sheng, Z. Xu, et al., Advantages of pure platelet-rich plasma compared with leukocyte and platelet-rich plasma in promoting repair of bone defects, *J. Transl. Med.* 14 (73) (2016).
- [5] B. Monteiro, R. Carlo, Association of mesenchymal stem cells with platelet-rich plasma on the repair of critical calvarial defects in mice, *Acta Cir. Bras. [Internet]* 27 (3) (2012) 1–11. Available from: [http://www.scielo.br/scielo.php?pid=S0102-86502012000300001&script=sci\\_arttext](http://www.scielo.br/scielo.php?pid=S0102-86502012000300001&script=sci_arttext).
- [6] J. Torres, Influence of platelet-rich plasma on bone regeneration: a histomorphometric study in rabbit calvaria, *J. Oral Maxillofac. Implant [Internet]* 22 (2007) 563–568. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/17929516>.
- [7] G. Spence, S. Phillips, C. Campion, R. Brooks, N. Rushton, Bone formation in a carbonate-substituted hydroxyapatite implant is inhibited by zoledronate, *J. Bone Jt. Surg.* 90 (8) (2008) 1635–1640.
- [8] M. Yoshioka, K. Tanimoto, Y. Tane, K. Sumi, T. Awada, N. Oki, et al., Bone regeneration in artificial jaw cleft by use of carbonated hydroxyapatite particles and mesenchymal stem cells derived from iliac bone [Internet], *Int. J. Dent.* (2012) Available from: <http://www.hindawi.com/journals/ijid/2012/352510/>.
- [9] L. Elhies, J. Carter, J. Natiella, J. Featherstone, D. Nelson, Quantitative analysis of early in vivo tissue response to synthetic apatite implants, *J. Biomed. Mater. Res.* 22 (2) (1988) 137–148.
- [10] N. Patel, J. Gibson, K. Hing, The in vivo response of phase pure hydroxyapatite and carbonate substituted hydroxyapatite granules of varying size ranges, *J. Key Eng. Mater.* 218 (2) (2002) 383–386.
- [11] M.R. Brinker, D. O'Connor, Basic sciences, in: A. Brigido (Ed.), Review of Orthopaedics, 5th ed., Elsevier, Philadelphia: Saunders, 2008, pp. 13–20.
- [12] V.J. Vigorita, Basic Science of Bone. Orthopaedic Pathology, 2nd ed., Lippincott Williams & Wilkins, 2000, pp. 9–41.
- [13] K. Schlegel, F. Kloss, P. Kessler, S.S. Mosgau, E. Nkenke, J. Wiltfang, Bone conditioning to enhance implant osseointegration: an experimental study in pigs, *Int. J. Oral Maxillofac. Implant.* 18 (1) (2003) 505–511.
- [14] T. Aghaloo, P. Moy, E. Freymiller, Investigation of platelet-rich plasma in rabbit cranial defects: a pilot study, *J. Oral Maxillofac. Surg.* 19 (1) (2004) 59–65.
- [15] M.E. Pryor, J. Yang, G. Polimeni, K.T. Koo, M.J. Hartman, H. Gross, et al., Analysis of rat calvaria defects implanted with a platelet-rich plasma preparation: radiographic observations, *J. Periodontol. Online* 76 (8) (2005) 1287–1292.
- [16] N. Broggin, W. Hofstetter, E. Hunziker, D. Bosshardt, M. Bornstein, I. Seto, et al., The influence of PRP on early bone formation in membrane protected defects: a histological and histomorphometric study in the rabbit calvaria, *Clin. Implant Dent. Relat. Res.* 13 (1) (2011) 1–12.